



## MINIMUM INHIBITORY CONCENTRATIONS OF CEPHALOSPORIN ANTIBIOTICS FOR FECAL AND URINARY *ESCHERICHIA COLI* FROM UNIVERSITY STUDENTS IN KEFFI, NIGERIA



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### Abstract

*Minimum inhibitory concentration (MIC) provides in-vitro justification for confirming antibiotic resistance strains. This study evaluated the MICs of cephalixin, cefuroxime, ceftazidime and cefepime for Escherichia coli isolated from urine and stool of university students in Keffi. Eighty bacteria (thirty from urine, fifty from stool) were isolated and identified as E. coli from students by cultural, microscopic and biochemical methods. MICs were evaluated using macro-broth dilution method and breakpoint susceptibility interpreted as described by Clinical and Laboratory Standards Institute (CLSI) of the United States of America (USA). The isolates were also screened for carriage of extended spectrum beta-lactamase (ESBL). The antibiotic susceptibility level and MIC for 50% of the isolates (MIC<sub>50</sub>) from urine was: cephalixin (0% and 45.8 µg/ml), cefuroxime (0% and 64.0 µg/ml), ceftazidime (20% and 18.4 µg/ml) and cefepime (47% and 13.8 µg/ml). Fecal isolates had susceptibility and MIC<sub>50</sub> as follows: cephalixin (8% and 34.2 µg/ml), cefuroxime (20% and 32.0 µg/ml), ceftazidime (32% and 8.0 µg/ml) and cefepime (46% and 4.8 µg/ml). With the exception of cefuroxime, the differences in MIC<sub>50</sub> values of each cephalosporin antibiotics tested for urine and stool isolates were insignificant. For both urinary and faecal isolates, MIC for 90% of the isolates (MIC<sub>90</sub>) decreased in the order: cefepime < ceftazidime < cefuroxime and cephalixin. This study revealed high susceptibility pattern of urinary and faecal E. coli isolates from apparently healthy individual to a fourth generation cephalosporin cefepime. Furthermore, all isolates are potential carriers of extended spectrum beta-lactamases. Further investigation is required to confirm the isolates as ESBL carriers.*

**Key Words:** Minimum Inhibitory Concentration; Cephalosporin; Antibiotics; *Escherichia coli*

### INTRODUCTION

Antimicrobial agents remain the mainstay treatment for infections by *Escherichia coli*, a common gastro-intestinal tract bacterium found in the large intestine of humans and other warm-blooded animals (Campbell & Reece, 2002) which is responsible for many intestinal and extra-intestinal infections (Bailey *et al.*, 2006). However, the continued usefulness of these agents is limited by the development of resistance mechanisms (Todar, 2007; Pitout & Laupland, 2008). Hence, the testing of the *in vitro* susceptibility of isolated bacteria to various antimicrobial agents is an important guide to antimicrobial therapy (Daivis & Dulbecco, 2000).

One important way to assess the antibiotic susceptibility of a bacterium is the determination of a “minimum inhibitory concentration” (MIC), the lowest

concentration of an antimicrobial that will inhibit the visible growth of the tested organism. The performance of antimicrobial susceptibility testing for bacterial isolates is an important task in the clinical microbiology laboratory that helps to detect possible drug resistance in common pathogens and assures susceptibility to drugs of choice for particular infections.

Resistance in *E. coli* isolates to cephalosporins is increasingly reported worldwide (Akins *et al.*, 2002; Forward *et al.*, 2004; Johnson *et al.*, 2007; Drawz & Bonomo, 2010; Ngwai *et al.*, 2010; Thokar *et al.*, 2010; Ngwai *et al.*, 2011; Park *et al.*, 2012; Talukdar *et al.*, 2013). It is thus of great concern to monitor the susceptibility of cephalosporins. This study evaluates the susceptibility of *E. coli* isolated from feces

and urine of university students in Keffi to cephalosporin antibiotics by MIC measurement. The goal of testing is to assure the susceptibility to cephalosporins of choice in *E. coli* infections and to detect possible resistance to cephalosporins. MICs are considered the 'gold standard' for determining the susceptibility of organisms to antimicrobials (Andrews, 2001).

## MATERIALS AND METHODS

### Bacterial Isolates

A total of 80 *E. coli* isolates (30 fecal, 50 urinary) were used in this study. They were isolated and identified from stool or urine of students of Nasarawa State University Keffi using standard cultural, microscopical and biochemical procedures (Cheesbrough, 2000). Pink colonies on MacConkey agar (BIOTEC Laboratories Ltd., Ipswich, United Kingdom) that grew with greenish metallic sheen characteristics on eosin methylene blue agar (BIOTEC Laboratories Ltd., Ipswich, United Kingdom) and which were indole positive, methyl red positive, Voges-Proskauer negative and citrate negative were confirmed as *E. coli*. Bacteria were stored in the refrigerator at 4°C on nutrient agar (NA: Merck KGaA, Darmstadt, Germany) slants and reactivated by sub-culturing on MacConkey agar and used for the experiments.

### Antibiotics

The antibiotics used were cephalixin (Ranbaxy Laboratories Ltd, India), cefuroxime (Glaxo Smith-Kline, India), ceftazidime (Glaxo Smith-Kline, Italy) and cefepime (Bharat Parenterals Ltd., India). All antibiotics were purchased from the Pharmacy Department, Federal Medical Center, Keffi, Nasarawa State, Nigeria. The stock solutions were prepared in appropriate solvents in accordance with the method of Clinical and Laboratory Standards Institute (CLSI, 2012).

### Determination of Minimum Inhibitory Concentration (MIC)

The MICs of the antibiotics against the *E. coli* isolates and quality control strain (*E. coli* ATCC 25922) were determined in triplicate using the CLSI macro-broth dilution method (CLSI, 2012). An adjusted inoculum of the test organism was inoculated into Mueller-Hinton broth (MHB: BIOTEC Laboratories Ltd., Ipswich, United Kingdom) containing two-fold dilutions of an initial antibiotic solution so that each tube contained approximately  $1 \times 10^5$  colony-forming units (CFU). Results were observed and registered after 24-h incubation at 37°C. MIC was defined as the lowest concentration that inhibited visible growth. Cumulative frequency curves of the antibiotic MICs of isolates were plotted and MICs for 50% (MIC<sub>50</sub>) and for 90% (MIC<sub>90</sub>) of isolates were then generated from the plots.

### Statistical Analyses

MIC<sub>50</sub> and MIC<sub>90</sub> were analyzed by one way analysis of variance (ANOVA) using Smith Statistical Package (SSP), version 2.80 (by Gary Smith, Pomona College, Claremont, California).

## RESULTS

### MIC ranges of antibiotics

The MIC ranges of the antibiotics for the *Escherichia coli* isolates are as shown in Table 1.

### MIC for 50% and 90% of isolates

The MIC for 50% of isolates (MIC<sub>50</sub>) and MIC for 90% of isolates (MIC<sub>90</sub>) generated from the cumulative frequency curves in Figure 1 (urinary) and Figure 2 (fecal) are as shown in Table 2.

### Susceptibility of isolates

The susceptibilities of the isolates to cephalosporin antibiotics are as shown in Table 3. The urinary isolates are slightly more resistant to the cephalosporins than

**Table 1:** Minimum Inhibitory Concentration ranges of Cephalosporin antibiotics for the *Escherichia coli* isolates

Antibiotics	MIC ranges (µg/ml)	
	Urinary	Fecal
Cephalexin	32-512	8-128
Cefuroxime	32-512	8-128
Ceftazidime	4-128	8-128
Cefepime	2-128	2-128

**Table 2:** Minimum Inhibitory Concentration of cephalosporin antibiotics for 50% and 90% of the *Escherichia coli* isolates

Antibiotics	MIC <sub>50</sub> (µg/ml)		MIC <sub>90</sub> (µg/ml)	
	Urinary	Fecal	Urinary	Fecal
Cephalexin	45.8 (≥ 4.0)	34.2 (≥ 4.0)	508.4 (≥ 4.0)	118.0 (≥ 4.0)
Cefuroxime	64.0 (≥ 4.0)	32.0 (≥ 4.0)	508.4 (≥ 4.0)	116.6 (≥ 4.0)
Ceftazidime	18.4 (≥ 16.0)	8.0 (≤ 16.0)	32.0 (≥ 16.0)	61.0 (≥ 16.0)
Cefepime	13.8 (≤ 16.0)	4.8 (≤ 16.0)	27.4 (≥ 16.0)	20.0 (≥ 16.0)

MIC<sub>50</sub> = Minimum Inhibitory Concentration for 50% of isolates; MIC<sub>90</sub> = Minimum Inhibitory Concentration for 90% of isolates.

the fecal isolates. Both isolate types were similarly susceptible to cefepime, a fourth generation cephalosporin antibiotic.

### Statistical Analyses

Antibiotic MIC<sub>50</sub> and MIC<sub>90</sub> for urine isolates were compared with those for fecal isolates. Significance of differences between the values were determined at p=0.05 (5% probability) as shown in Table 4. The statistical analyses indicated that the antibiotics have insignificant p-values; while the comparison of the sources of the isolates was be significant.

### DISCUSSION

The testing of the *in vitro* susceptibility of isolated bacteria to various antimicrobial agents is an important guide to therapy (Daivis & Dulbecco, 2000). This is because the susceptibility of bacterial strains to a given antibiotic can grossly affect the *in vivo* efficacy of that antibiotic, among other factors like age, immune status of patient, existing disorder and route of antibiotic administration (Mandell, 2002). The determination of MIC is one important way to assess the antibiotic

susceptibility of a bacterium. Minimum inhibitory concentration values are used in diagnostic laboratories, mainly to confirm resistance of strains of bacteria (Andrew, 2001). This study evaluated the minimum inhibitory concentration of cephalosporin antibiotics against *E. coli* from fecal and urinary sources among university students in Keffi.

With MIC values obtained for some of the isolates above the CLSI resistance breakpoint of ≥4 µg/ml (cephalexin and cefuroxime), ≥16 µg/ml (ceftazidime) and ≥32 µg/ml cefepime) (CLSI, 2010), such isolates are said to be resistant to the cephalosporin antibiotics tested. Previous studies on *E. coli* elsewhere have reported cephalosporin resistance (Diekema *et al.*, 2000; Feng *et al.*, 2002; Ehinmidu, 2003; Ngwai *et al.*, 2005; Zing *et al.*, 2006; Ngwai *et al.*, 2010; Ngwai *et al.*, 2011; Park *et al.*, 2012; Talukdar *et al.*, 2013).

Since MIC values of ceftazidime obtained for 50% and 90% of isolates at ≥2 µg/ml, these isolates are potential carriers of extended spectrum beta-lactamases (ESBL) since each *E.*

*coli*, *Klebsiella pneumoniae* or *K. oxytoca* isolate is considered a potential ESBL producer if MIC test results show  $\geq 2$   $\mu\text{g/ml}$  for at least one of the extended-spectrum cephalosporins (cefepodoxime, ceftazidime, ceftriaxone or cefotaxime) and monobactams (e.g. aztreonam) (NCCLS, 1999). The possibility of our isolates being ESBL carriers is justified by the fact that studies using cefepodoxime or ceftazidime show the highest sensitivity for ESBL detection (Ho *et al.*, 2000; MacKenzie *et al.*, 2002). Many clinical and non-clinical *E. coli* isolates are known to produce extended spectrum beta-

lactamases (ESBL) which are plasmid borne (Paterson & Bonomo, 2005; Pitout & Laupland, 2008; Isendahl *et al.*, 2012; Al-Mayahie, 2013).

The generally higher susceptibility observed for the *E. coli* isolates to cefepime is not new (Iqbal *et al.*, 2002); and should not be taken for granted to suggest clinical usefulness in view of CLSI's recommendation that all cephalosporin susceptibility results be reported as "resistant" when an isolate is determined to produce ESBL (CLSI, 2009).

**Table 3:** Susceptibility of *Escherichia coli* isolates to cephalosporin antibiotics

Antibiotics	Number (%) Susceptibility	
	Urinary (n = 30)	Fecal (n = 50)
Cephalexin	0 (0)	4 (8)
Cefuroxime	0 (0)	10 (20)
Ceftazidime	6 (20)	16 (32)
Cefepime	14 (47)	23 (46)

**Table 4:** Statistical analyses of Minimum Inhibitory Concentration of Cephalosporin antibiotics for 50% and 90% of *Escherichia coli* isolates

Statistics	Isolate Type	P value	Remarks (at P = 0.05)
MIC <sub>50</sub> Cephalexin	Urine vs Stool	0.4226	Insignificant
MIC <sub>50</sub> Cefuroxime	Urine vs Stool	0.0073	Significant
MIC <sub>50</sub> Ceftazidime	Urine vs Stool	0.1703	Insignificant
MIC <sub>50</sub> Cefepime	Urine vs Stool	0.5206	Insignificant
MIC <sub>90</sub> Cephalexin	Urine vs Stool	0.0029	Significant
MIC <sub>90</sub> Cefuroxime	Urine vs Stool	0.0029	Significant
MIC <sub>90</sub> Ceftazidime	Urine vs Stool	0.1358	Insignificant
MIC <sub>90</sub> Cefepime	Urine vs Stool	0.0002	Significant

MIC<sub>50</sub> = Minimum Inhibitory Concentration for 50% of isolates; MIC<sub>90</sub> = Minimum Inhibitory Concentration for 90% of isolates.

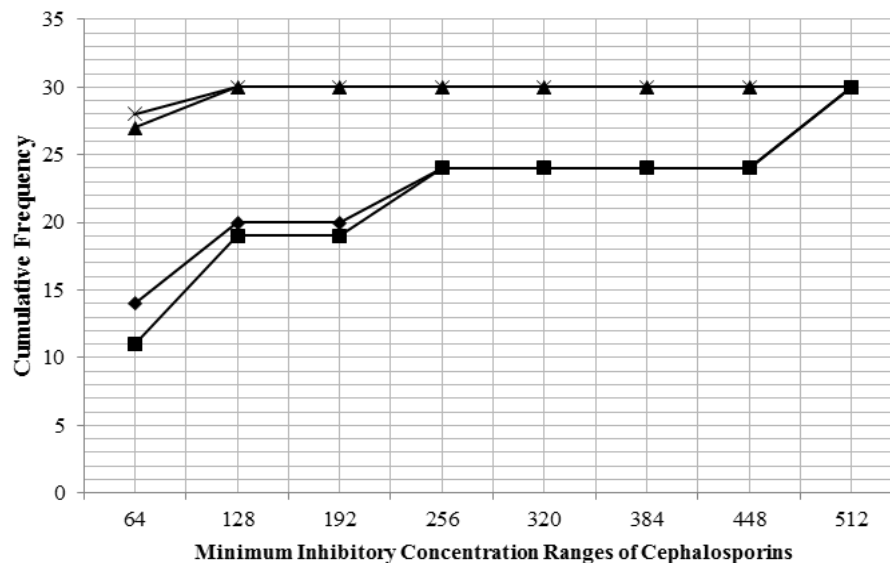
The result from this study provides possible evidence for higher susceptibility, to the fourth generation cephalosporin cefepime, by *E. coli* isolated from university students in Keffi decreasing in the order: cefepime < ceftazidime < cefuroxime < cephalexin. Furthermore, all isolates are potential carriers of extended spectrum beta-lactamases. Further investigation

is required to confirm the isolates as ESBL carriers.

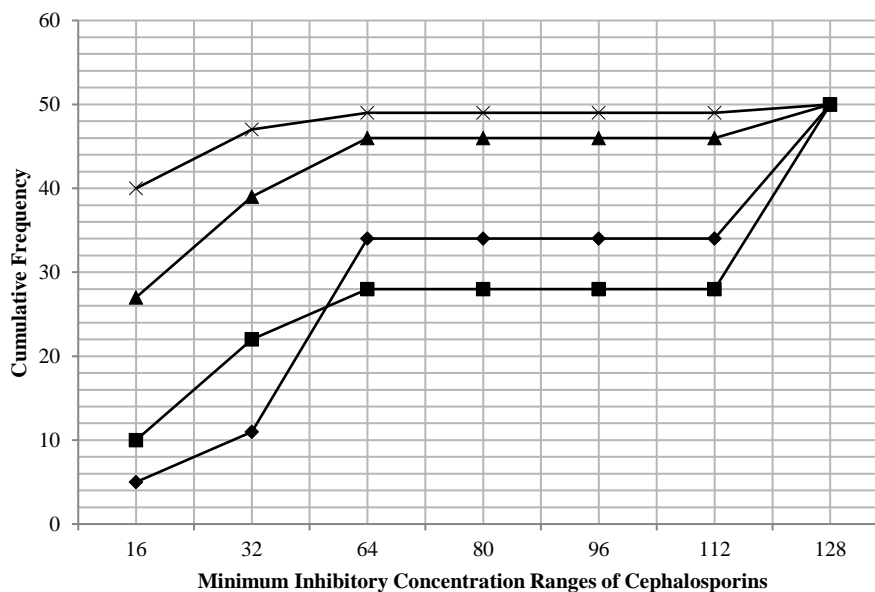
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*Minimum inhibitory concentrations of cephalosporin antibiotics for fecal and urinary Escherichia coli from university students in Keffi, Nigeria*



**Figure 1:** Cumulative Frequency Curves of Cephalosporin MICs for urinary isolates of *Escherichia coli* (◆—Cephalexin, ■—Cefuroxime, ▲—Ceftazidime, X—Cefepime).



**Figure 2:** Cumulative Frequency Curves of Cephalosporin MICs for fecal isolates of *Escherichia coli* (Cephalexin ◆, Cefuroxime ■, Ceftazidime ▲, Cefepime X).

## REFERENCES

- Akins, R.L., Haase, K.K. & Morris, A.J. (2002). Comparison of various fluoroquinolones (FQs) and four other antibiotics by mutant prevention concentration (MPC) against multi-drug resistant gram-negatives utilizing kill curves based on MPC-derived doses. In: Program and Abstracts of the Forty-second Interscience Conference on Antimicrobial Agents and Chemotherapy, San Diego, CA. A-1211, p. 10. American Society for Microbiology, Washington, DC, USA.
- Al-Mayahie, S.M.G. (2013). Phenotypic and genotypic comparison of ESBL production by Vaginal *Escherichia coli* isolates from pregnant and non-pregnant women. *Ann. Clin. Microbiol. Antimicrob.*, 12: 7.
- Andrews, J. M. (2001). Determination of Minimum Inhibitory Concentrations. *J. Chemother.*, 48(Suppl.1): 5-6.
- Bailey, M.T., Engler, H. & Sheridan, J.F. (2006). The translocation of cutaneous and gastrointestinal microflora to secondary lymphoid organs of C57BL/6 mice. *J. Neuroimmunol.*, 171: 29-37.
- Campbell, N.A. & Reece, J.B. (2002). *Biology*. Pearson Education Inc: San Francisco.
- Cheesbrough, M. (2000). *District Laboratory practice in Tropical Countries*. Part 2. Cambridge University Press: United Kingdom. pp 63-70.
- CLSI. (2009). Performance standards for antimicrobial susceptibility testing. Nineteenth Informational Supplement. Document M100-S19. Clinical and Laboratory Standards Institute: Wayne, Pennsylvania.
- CLSI. (2010). Recommended breakpoint changes for selected cephalosporins and aztreonam for Enterobacteriaceae. M100-S20. Clinical and Laboratory Standards Institute: Wayne, PA.
- CLSI. (2012). *Methods for Dilution Susceptibility Tests for Bacteria That Grow Aerobically*; Approved Standard- Ninth Edition (M07-A9). Clinical and Laboratory Standards Institute: Wayne, PA.
- Daivis, B.D. & Dulbecco, R. (2000). *Microbiology: General aspects of chemotherapeutic action*. 5th ed. J.B. Lippincott Company: Philadelphia.
- Diekema, D.J., Pfaller, M.A., Jones, R.N., Doern, G.V., Kugler, K.C., Beach, M.L., Sader, H.S. & The SENTRY Participants Group. (2000). Trends in antimicrobial susceptibility of bacterial pathogens isolated from patients with bloodstream infections in the USA, Canada and Latin America. *Int. J. Antimicrob. Agents* 13: 257-271.
- Drawz, S.M. & Bonomo, R.A. (2010). Three decades of beta-lactamase inhibitors. *Clin. Microbiol. Rev.*, 23: 160-201.
- Ehinmidu, J.O. (2003). Antibiotics susceptibility patterns of urine bacterial isolates in Zaria, Nigeria. *Trop. J. Pharm. Res.*, 2(2): 223-228.
- Feng, P., Weagant, S., Grant, M. (2002). Enumeration of *Escherichia coli* and Coliform Bacteria. *Bacteriological Analytical Manual*. 8th ed. FDA/Center for Food Safety and Applied Nutrition. <http://www.cfsa.fda.gov/Ebam/bam.e.html>.
- Forward, K.R., Matheson, K.M., Hiltz, M., Musgrave, H. & Poppe, C. (2004). Recovery of cephalosporin-resistant *Escherichia coli* and *Salmonella* from pork, beef and chicken marketed in Nova Scotia. *Can. J. Infect. Dis. Med. Microbiol.*, 15(4): 226-230.
- Ho, P.L., Tsang, D.N., Ho, M. & Yuen, K.Y. (2000). Comparison of screening methods for detection of extended spectrum beta-lactamases and their prevalence among *Escherichia coli*

- and *Klebsiella* species in Hong Kong. APIMS 108: 237-240.
- Iqbal, M., Patel IK, Ain, Q., Barney, N., Kiani, Q., Rabbani, K.Z., Zaidi, G., Mehdi, B., Shah, S.H. (2002). Susceptibility Patterns of *Escherichia coli*: Prevalence of Multidrug-resistant Isolates and Extended Spectrum Beta-Lactamase Phenotype. J. Pak. Med. Assoc., 52: 407.
- Isendahl, J., Turlej-Rogacka, A., Manjuba, C., Rodrigues, A., Giske, C.G. & Nauc  r, P. (2012). Fecal Carriage of ESBL-Producing *E. coli* and *K. pneumoniae* in Children in Guinea-Bissau: A Hospital-Based Cross-Sectional Study. PLoS One 7(12): e51981. doi:10.1371/journal.pone.0051981.
- Johnson, J.R., Sannes, M.R., Croy, C., Johnston, B., Clabots, C., Kuskowski, M.A., Bender, J., Smith, K.E., Winokur, P.L. & Belongia, E.A. (2007). Antimicrobial Drug-Resistant *Escherichia coli* from humans and poultry products, Minnesota and Wisconsin, 2002-2004. Emerg. Infect. Dis., 13 (6). <http://wwwnc.cdc.gov/eid/article/13/6/06-1576.htm>. Accessed Date: 15/05/2013.
- MacKenzie, F.M., Miller, C.A. & Gould, I.M. (2002). Comparison of screening methods for TEM- and SHV-derived extended spectrum beta-lactamase detection. Clin. Microbiol. Infect., 8: 715-24.
- Mandell, G.L. (2002). Principles and practice of infectious disease: Anti-infective therapy. 4th ed. Churchill Livingstone: New York.
- NCCLS. (1999). Performance standards for antimicrobial susceptibility testing: approved standard. M100-S9. National Committee for Clinical Laboratory Standards: Wayne (PA).
- Ngwai YB, Akpotu MO, Obidake RE, Sounyo AA, Onanuga A, Origbo SO (2010). Antimicrobial Susceptibility of *Escherichia coli* and other coliforms isolated from urine of asymptomatic students in Bayelsa State, Nigeria. Afr. J. Microbiol. Res. 5(3): 184-191.
- Ngwai, Y.B., Ezenwa, F.C. & Ngadda, N. (2011). Contamination of Nigerian currency notes by *Escherichia coli* in Nasarawa State University, Keffi, Nigeria. Asian J. Pharm. Hea. Sci., 1(4): 163-166.
- Ngwai, Y.B., Onaolapo, J.A., Ehinmidu, J.O., Ibrahim, Y.K.E. & Olutimayin, G. (2005). Frequency of ampicillin resistance amongst uropathogenic strains of *Escherichia coli* isolated from patients with suspected urinary tract infections in Zaria. Nigerian J. Pharm. Sci., 6: 26-32.
- Park, Y.S., Adams-Haduch, J.M., Shutt, K.A., Yarabinec III, D.M., Johnson, L.E., Hingwe, A., Lewis II, J.S., Jorgensen, J.H. & Doi, Y. (2012). Clinical and Microbiologic Characteristics of Cephalosporin-Resistant *Escherichia coli* at Three Centers in the United States. Antimicrob. Agents Chemother. 56 (4): 1870-1876.
- Paterson, D.L. & Bonomo, R.A. (2005). Extended-spectrum beta-lactamases: a clinical update. Clin. Microbiol. Rev., 18: 657-686.
- Pitout, J.D. & Laupland, K.B. (2008). Extended-spectrum beta-lactamase-producing Enterobacteriaceae: an emerging public-health concern. Lancet Infect. Dis., 8: 159-166.
- Talukdar, P.K., Rahman, M., Rahman, M., Nabi, A., Islam, Z., Hoque, M.M., Endtz, H.P. & Islam, M.A. (2013). Antimicrobial Resistance, Virulence Factors and Genetic Diversity of *Escherichia coli* Isolates from Household Water Supply in Dhaka, Bangladesh. PLoS ONE 8(4): e61090. doi:10.1371/journal.pone.0061090.
- Thokar, M.A., Fomda, B.A., Maroof, P., Ahmed, K., Bashir, D. & Bashir, G. (2010). Proliferation of extended spectrum beta-Lactamase (ESBL) producing gram negative bacteria,

- diagnostic inputs and impact on selection of antimicrobial therapy. Physicians Acad., 4: 25-31.
- Todar, K. (2007). "Pathogenic *Escherichia coli*." Online Textbook of Bacteriology. University of Wisconsin-Madison. Department of Bacteriology.
- <http://www.textbookofbacteriology.net/e.coli.html>. Accessed Date: 05/03/2013.
- Zing, G.F., Jase, K.S. & Hugh, N.S. (2006). Bacterial Plurality as a General Mechanism Driving Persistence in Chronic Infections. Clin. Chemother., 437: 20-24.